Impact of tree choice in metagenomics differential abundance studies

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Work and slides done by Antoine Bichat during his Ph.D., cosupervised by C. Ambroise and J. Plassais

Context

Microbiota

Ecological community of microorganisms that reside in an environmental niche

Some figures for human gut

- 10¹⁴ bacterial cells in one gut...
- ... weighing 2 kg
- More than 1500 different species
- More than 10 millions unique genes



Proven associations

- Immune system
- Crohn's disease
- Vaginosis
- Diabete
- Tobacco
- Diet
- Antibiotics
- Birth mode



Gut: The Inside Story of Our Body's Most Underrated Organ (Giulia Enders)

Data - abundances of taxa

Warning: The `n_extra` argument of `print()` is deprecated as of pillar 1.6.2. Please use the `max_extra_cols` argument instead.

A tibble: 122 × 395 S001 S002 S003 S005 S006 S007 S008 Таха S004 S009 S010 <chr> <dbl> <dbl > 1 Lactob... 2318 2 Prevot. 3 Megasp... 4 Sneath... 5 Atopob... 6 Strept... 7 Dialis... 8 Anaero... 9 Pepton... 10 Eggert...

... with 112 more rows

- Count data (or compositional) data
- Zero-inflated data
- Correlation between species

• Counts spanning several orders of magnitude: $1 \rightarrow 10^8$ Ravel et al. (2011)



Data - taxonomy

#	A tibble: 129 × 5				
	Phylum	Class	Order	Family	Genus
	<chr></chr>	<chr></chr>	<chr></chr>	<chr></chr>	<chr></chr>
1	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Actinobaculum
2	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Actinomyces
3	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Arcanobacteri
4	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Mobiluncus
5	Actinobacteria	Actinobacteria	Actinomycetales	Actinomycetaceae	Varibaculum
#	with 124 more	e rows			

Aerococcus Aerococcacea Facklamia Lactococcus Streptococcacea Streptococcus Lactobacilla Enterococcus Lactobacillus Bacil Exiguobacterium Bacillales_incertae_sedi Gemella Bacillale Staphylococcus Parvimonas Peptoniphilus Peptoniphilacea Anaerococcus Finegoldia Dialister Firmicute Anaeroglobus Veillonellacea Megasphaera Veillonella Peptostreptococcus Clostridiale Peptococcus Moryella Bulleidia Arcanobacterium Actinomyces Actinomycetacea Mobiluncus Varibaculum Corynebacterium Corynebacteriale Actinobacte Segniliparus Propionibacterium Actinobacteri Gardnerella Eggerthella Coriobacterii Atopobium Bacter Bacteroides Bacteroidal Prevotella Porphyromonas Campylobacter Proteobacteri Stenotrophomonas Sutterella Fusobacterium Fusobacteriale Sneathia Ureaplasma

Differential abundance studies

Evaluation of binary classifiers

The true condition is usually unknown for real dataset.

The prediction is usually determined by comparing the *p*-value to $\alpha = 0.05$.



Statistical issue

Univariate tests on hundred of taxa

Need for a multiple testing controling procedure!

Can we use it to do it better?

- Hierarchical FDR
- *z*-scores smoothing



Philippot et al. (2010)



Hierarchical FDR

This procedure inscreases statistical power by lessening the number of test to do with a descending method:

- Test the family \mathcal{T}_0
- If node t is rejected, test $\mathcal{T}_t = \{H_i \mid \operatorname{Par}(i) = t\}$ with a BH procedure at level q



This procedure controls the FDR at level

 $1.44 imes q imes rac{\# \mathrm{discoveries} + \# \mathrm{families \ tested}}{\# \mathrm{discoveries} + 1}$



z-scores smoothing

Denote by **z** the vector of observed *z*-scores and μ the vector of "true" *z*-scores

Assume that $\mathbf{z}|\mu \sim \mathcal{N}_n\left(\mu,\sigma^2\mathbf{I}_m
ight)$ and $\mu \sim \mathcal{N}_m\left(\gamma\mathbf{1},\tau^2\mathbf{C}_
ho
ight)$

then

$$\mathbf{z} \sim \mathcal{N}_m\left(\gamma \mathbf{1}, au^2 \mathbf{C}_
ho + \sigma^2 \mathbf{I}_m
ight)$$

and Bayes formula gives

$$\mu^* = \left(\mathbf{I}_m + rac{\sigma_0^2}{ au_0^2} \mathbf{C}_{
ho_0}^{-1}
ight)^{-1} \left(rac{\sigma_0^2}{ au_0^2} \mathbf{C}_{
ho_0}^{-1} \gamma_0 \mathbf{1} + \mathbf{z}
ight)^{-1}$$

with σ_0 , τ_0 , ρ_0 and γ_0 hypermarameters

After smoothing, a multiple testing correction could be done on smoothed values



Which tree?

Taxonomy? Phylogeny?

- Proxy for correlations at high-level niches
- Not so much for low-level niches?
- Not available everytime

Correlation tree?

- Actual correlation between taxa
- Computed from data using pairwise correlation



Comparison of trees

Billera-Holmes-Vogtamnn distance

The BHV distance is the length of the unique shortest path between the trees on treespace







Quantifying distance between trees

- trees of primary interest
 - correlation tree on original data
 - taxonomy
- what is the confident region for the correlation tree?
 - correlation trees on boostrapped data (resampling on samples)
- are trees significantly closer than two random trees?
 - trees created by random shuffling of correlation tree tip labels
 - trees created by random shuffling of taxonomy tip labels

We compute all pairwise distances between these trees



Random shuffling





Dataset

- Vaginal microbiome of non pregnant women sequenced by 16S
- 40 different genera after filtering (~30%)





Correlation tree

Taxonomy



15 / 35

📄 Ravel et al. (2011)

Pairwise distances



😀 The correlation tree is different from the taxonomy



Evaluation of hFDR

Chlamydiae dataset

- Small subset of the GlobalPatterns dataset narrowed to Chlamydiae phylum
- 21 different OTUs
- 26 samples representing 9 very different environments: soil, ocean, feces, skin...

Method

- Find which bacteria are differentially abundant between environments
- Association using Fisher statistic (ANOVA)
- Correction with hierarchical FDR

Abundances of detected species





Representation of evidences on trees



Correlation correction



But...

 $\alpha = 0.10$ is only the family-level FDR.

The *a posteriori* global FDR is:

- $\alpha' = 0.32$ for phylogenetic correction
- $\alpha' = 0.324$ for correlation correction

A BH procedure at the same global FDR level leads to 15 discoveries (+5)

😀 Using correlation tree instead of taxonomy yields more results

🙁 Vanilla BH beats hFDR for a given level



Chaillou Dataset

- Food-associated microbiota of processed meat and seafood products
- 97 different OTUs
- 80 samples across 8 different food type: beef, veal, salmon, shrimp...



Abundances of detected species





Impact of the test



Position of selected OTUs on the correlation tree



OTUs in red are only detected by phylogeny

All OTU in the clade are differentially abundant

Obvious unequal variances between groups

KW test is more robust than F-test in this setting

Implemented tests are not appropriate to metagenomic data



Evaluation of z-scores smoothing

Zeller dataset

- Dataset from cancer study
- 119 different genera (after filtering)
- 199 samples: 42 adenoma, 91 carcinoma and 66 control

Method

- Find which bacteria are differentially abundant between diseases
- Association using Kruskal-Wallis test
- Correction with hierarchical *p*-value smoothing



Impact of the tree



- 😊 z-scores smoothing is slightly better than vanilla BH
- 🙁 All hierachies give highly similar results



Simulations

Workflow

- Simulate DA taxa starting from an homogeneous dataset
- Correction with BH and hierarchical *p*-value smoothing







Change in z-scores

30 / 35

 $\mathrm{Mean} \ \mathrm{z}\mathrm{-smoothing} = |z_\mathrm{raw} - z_\mathrm{adjusted}|$



🙁 Taxonomy behaves like random tree and has little impact

In most cases, smoothing has absolutely no impact on the result



Evaluation



😀 Using correlation tree instead of taxonomy yields more results

- 😕 Vanilla BH is better
- Taxonomy is worse than random trees



Conclusions

Conclusions

Correlation tree and taxonomy are very different Replacing taxonomy tree with correlation tree increases the TPR

🙁 Vanilla BH is more powerful than hFDR

Bayesian smoothing does not really depend on the tree for z-scores smoothing Overall incorporating phylogenetic is not tremendously helpful...



References

- Bichat, A. et al. "Incorporating Phylogenetic Information in Microbiome Differential Abundance Studies Has No Effect on Detection Power and FDR Control" Frontiers in Microbiology 11:649 (2020). doi: 10.3389/fmicb.2020.00649
- Blander, J. Magarian, et al. "Regulation of inflammation by microbiota interactions with the host." Nature immunology 18.8 (2017): 851.
- Morgan, Xochitl C., et al. "Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment." Genome biology 13.9 (2012): R79.
- Ravel, Jacques, et al. "Vaginal microbiome of reproductive-age women." Proceedings of the National Academy of Sciences 108.Supplement 1 (2011): 4680-4687.
- Qin, Junjie, et al. "A metagenome-wide association study of gut microbiota in type 2 diabetes." Nature 490.7418 (2012): 55.
- Opstelten, Jorrit L., et al. "Gut microbial diversity is reduced in smokers with Crohn's disease." Inflammatory bowel diseases 22.9 (2016): 2070-2077.
- Bokulich, Nicholas A., et al. "Antibiotics, birth mode, and diet shape microbiome maturation during early life." Science translational medicine 8.343 (2016): 343ra82-343ra82.



Thanks for your attention!

Questions?